

Predicting Maize Phenology: Intercomparison of Functions for Developmental Response to Temperature

S. Kumudini, F. H. Andrade, K. J. Boote, G. A. Brown, K. A. Dzotsi, G. O. Edmeades, T. Gocken, M. Goodwin, A. L. Halter, G. L. Hammer, J. L. Hatfield, J. W. Jones, A. R. Kemanian, S.-H. Kim, J. Kiniry, J. I. Lizaso, C. Nendel, R. L. Nielsen, B. Parent, C. O. Stöckle, F. Tardieu, P. R. Thomison, D. J. Timlin, T. J. Vyn, D. Wallach, H. S. Yang, and M. Tollenaar*

ABSTRACT

Accurate prediction of phenological development in maize (*Zea mays* L.) is fundamental to determining crop adaptation and yield potential. A number of thermal functions are used in crop models, but their relative precision in predicting maize development has not been quantified. The objectives of this study were (i) to evaluate the precision of eight thermal functions, (ii) to assess the effects of source data on the ability to differentiate among thermal functions, and (iii) to attribute the precision of thermal functions to their response across various temperature ranges. Data sets used in this study represent >1000 distinct maize hybrids, >50 geographic locations, and multiple planting dates and years. Thermal functions and calendar days were evaluated and grouped based on their temperature response and derivation as empirical linear, empirical nonlinear, and process-based functions. Precision in predicting phase durations from planting to anthesis or silking and from silking to physiological maturity was evaluated. Large data sets enabled increased differentiation of thermal functions, even when smaller data sets contained orthogonal, multi-location and -year data. At the highest level of differentiation, precision of thermal functions was in the order calendar days < empirical linear < process based < empirical nonlinear. Precision was associated with relatively low temperature sensitivity across the 10 to 26°C range. In contrast to other thermal functions, process-based functions were derived using supra-optimal temperatures, and consequently, they may better represent the developmental response of maize to supra-optimal temperatures. Supra-optimal temperatures could be more prevalent under future climate-change scenarios, but data sets in this study contained few data in that range.

Modeling crop productivity under climate change has been a topic of much discussion, and maize is particularly of significance due to its importance as a global source of food and feed. Model predictions of crop productivity under climate change are known to be hindered by uncertainties inherent in these analyses. Uncertainties can arise from a number of sources including the choice of general circulation model, the downscaling climate methodology, as well as variation among crop models. In a recent report, Asseng et al. (2013) found that a greater proportion of the uncertainty in simulating the response of crop yields to climate change is attributable to differences in crop models than the uncertainties due to variations among down-scaled general circulation models. Crop model differences can arise from variations in crop parameters or the approaches used to simulate the underlying biophysical processes. There is a need to evaluate the accuracy of the processes underlying crop models, specifically those processes that are most immediately affected by the environmental factors modified by climate change, such as temperature, water, and elevated CO₂.

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The impact of increased temperature due to climate change on crop production has been a topic of much interest in the scientific literature. The Intergovernmental Panel on Climate Change forecast an increase in global average temperature of 1.8 to 4°C from the period 1980 to 1999 to the period 2080

S. Kumudini and M. Tollenaar, The Climate Corp., 110 T.W. Alexander Drive, Research Triangle Park, NC 27709; F.H. Andrade, Unidad Integrada INTA EEA Balcarce-Facultad de Ciencias Agrarias, Univ. Nacional de Mar del Plata, C.C. 276 (7620) Balcarce, Argentina; K.J. Boote, Dep. of Agronomy, P.O. Box 110500, Univ. of Florida, Gainesville, FL 32611; G.A. Brown, Breaking Ground, 3501 W. Fillmore St, Chicago, IL 60624; K.A. Dzotsi and J.W. Jones, Dep. of Agricultural and Biological Engineering, P.O. Box 110570, Univ. of Florida, Gainesville, FL 32611; G.O. Edmeades, 43 Hemans St., Cambridge 3432, New Zealand; T. Gocken and M. Goodwin, Monsanto Co., St Louis, MO; A.L. Halter, Dupont-Pioneer, 5153 E. Simpson Dr., Vincennes, IN 47591; G.L. Hammer, QAAFI, Univ. of Queensland, Brisbane, QLD 4072, Australia; J.L. Hatfield, USDA-ARS, National Lab. for Agriculture and Environment, Ames, IA 50011; A.R. Kemanian, Dep. of Plant Science, Pennsylvania State Univ., University Park, PA 16802; S.-H. Kim, School of Environmental and Forest Sciences, College of the Environment, Univ. of Washington, Seattle, WA 98195; J. Kiniry, USDA-ARS, 808 E. Blackland Rd., Temple, TX 76502; J.I. Lizaso, Dep. Producción Vegetal: Fitotecnia, Univ. Politécnica de Madrid, 28040 Madrid, Spain; C. Nendel, Institute of Landscape Systems Analysis, Leibniz Centre for Agricultural Landscape Research, 15374 Müncheberg, Germany; R.L. Nielsen and T.J. Vyn, Dep. of Agronomy, Purdue Univ., West Lafayette, IN 47907; B. Parent and F. Tardieu, INRA, UMR759 Lab. d'Ecophysiologie des Plantes sous Stress Environnementaux, Place Viala, F-34060 Montpellier, France; C.O. Stöckle, Biological Systems Engineering, Washington State Univ., Pullman, WA 99164; P.R. Thomison, Dep. of Horticulture and Crop Science, Ohio State Univ., Columbus, OH 43210; D.J. Timlin, USDA-ARS, Crop Systems and Global Change Lab., Beltsville, MD 20705; D. Wallach, INRA, UMR 1248 Agrosystèmes et développement territorial (AGIR), 31326 Castanet-Tolosan Cedex, France; and H.S. Yang, Dep. Agronomy and Horticulture, Univ. of Nebraska, Lincoln, NE 68583. Received 14 Apr. 2014. *Corresponding author (matthijs.tollenaar@climate.com).

Abbreviations: CHU, crop heat units; GTI, general thermal index; GDD, growing degree days; RM, relative maturity; TLU, thermal leaf units.

Table 1. Summary of thermal functions, input temperatures (daily), temperature boundary conditions, and instantaneous and daily rates of thermal accumulation. The general thermal index (GTI) model has separate functions for the pre- and post-flowering periods (GTIveg and GTIrep, respectively).

Model type	Model name	Input temperatures (T)†		Boundary conditions		Thermal accumulation		Reference
		T _{min} and T _{max}	T (1 h)	T _{min} < 4.4°C then T _{min} = 4.4°C T _{min} < 10°C then T _{max} = 10°C	T > 34°C then T = 34°C	Instantaneous rate (IR)	Daily rate	
Empirical linear	GDD _{10,30}	T _{min} and T _{max}	T (1 h)	T < 10°C then T = 10°C T > 30°C then T = 30°C	T > 34°C then T = 34°C	$(T_{\min} + T_{\max})/2 - 10^{\circ}\text{C}$	IR	Gilmore and Rogers (1958)
	CERES-Maize	T _{min} and T _{max}	T (1 h)	T < 8°C then T = 8°C T > 34°C then T = 34°C		T - 8°C	$\Sigma(\text{IR}/24)$	Jones and Kiniry (1986)
Empirical nonlinear	CHU	T _{min} and T _{max}	T _{min} and T _{max}	T _{min} < 4.4°C then T _{min} = 4.4°C T _{min} < 10°C then T _{max} = 10°C		$\text{CHU}_{\min} = 1.8(T_{\min} - 4.4^{\circ}\text{C})$ $\text{CHU}_{\max} = 3.33(T_{\max} - 10^{\circ}\text{C}) - 0.084(T_{\max} - 10^{\circ}\text{C})^2$	$(\text{CHU}_{\max} + \text{CHU}_{\min})/2$	Brown and Bootsma (1993)
	GTIveg GTIrep	T _{mean} T _{mean}	T _{mean} T _{mean}			$0.043177T^2 - 0.000894T^3$ $5.3581 + 0.011178T^2$	IR IR	Stewart et al. (1998) Stewart et al. (1998)
Process based	APSIM	T (3 h)	T (3 h)	T < 0°C then T = 0°C T > 44°C then T = 44°C		$0^{\circ}\text{C} = < T < 10^{\circ}\text{C}; \text{IR} = T(10/18^{\circ}\text{C})$ $18^{\circ}\text{C} = < T < 34^{\circ}\text{C}; \text{IR} = T - 8^{\circ}\text{C}$ $34^{\circ}\text{C} = < T < 44^{\circ}\text{C}; \text{IR} = 26^{\circ}\text{C} - (T - 34^{\circ}\text{C})/2.6$	$\Sigma(\text{IR}/8)$	Wilson et al. (1995)
	TLU	T _{min} and T _{max}	T _{min} and T _{max}	T < 6°C then T = 6°C T > 44.8°C then T = 44.8°C		$\text{TLU}_{\min} = 0.0997 - 0.036T_{\min} + 0.00362T_{\min}^2$ $-0.000639T_{\min}^3$ $\text{TLU}_{\max} = 0.0997 - 0.036T_{\max} + 0.00362T_{\max}^2$ $-0.000639T_{\max}^3$	$(\text{TLU}_{\max} + \text{TLU}_{\min})/2$	Tollenaar et al. (1979)
	MAIZSIM	T (1 h)	T (1 h)	T < 0°C then T = 0°C T > 43.7°C then T = 43.7°C		$0.53[(43.7 - T)/(11.6)(T/32.1)]^{2.767}$	$\Sigma(\text{IR}/24)$	Kim et al. (2012)
	EnzymResp	T (1 h)	T (1 h)			$(T + 273)\exp\{-73900/[8.314(T + 273)]\}/[1 + (\exp\{-73900/[8.314(T + 273)]\})^{3.5}][1 - (T + 273)/306.4]$	$\Sigma(\text{IR}/24)$	Parent and Tardieu (2012)

† T_{min}, daily minimum temperature; T_{max}, daily maximum temperature; T(1 h), estimated from daily T_{min} and T_{max} using sine-wave temperature distribution during a 24-h period (e.g., Tollenaar et al., 1979); T_{mean}, daily mean temperature; T(3 h), estimated from range fractions that describe a sine wave.

to 2099 (Intergovernmental Panel on Climate Change, 2007). Temperature increases can impact crop production in a number of ways, but arguably the most important of these is the impact of temperature on crop phenology. The importance of phenology for crop productivity is well understood. The phenology of a crop will determine its adaptation to a region, its ability to mature and set grain within a growing season, and the synchrony of key developmental phases with ambient environmental conditions critical for productivity. Consequently, differences in model prediction of crop developmental phases can have significant impacts on the accuracy of the effects of forecast climate change on crop productivity.

There are a variety of thermal functions in current crop models that capture the relationship between temperature and maize development. These functions vary in a number of ways, including parameter values, the quantity and quality of the data from which they were formulated, the complexity of the algorithm used to capture the relationship, and the manner in which the algorithms were derived (i.e., empirical or process based) (Table 1). The relationship between temperature and maize development can be derived either empirically or through a process-based methodology. For the purposes of this study, empirically derived functions are defined as those that are formulated and parameterized utilizing the same measured phenomenon as the phenomenon to be predicted (e.g., time from planting to anthesis). Thermal functions derived through process-based methodology are defined as those functions that are parameterized and derived utilizing information at a lower level of organization than the phenomenon that is to be predicted (e.g., enzyme kinetics to predict the time from planting to anthesis). The thermal functions studied here are grouped into three function types that represent simple linear empirical relationships (Gilmore and Rogers, 1958; Jones and Kiniry, 1986), more complex empirical nonlinear relationships (Brown, 1969; Stewart et al., 1998), and functions based on process-based relationships between temperature and biochemical or organ-level responses (Tollenaar et al., 1979; Kim et al., 2012; Parent and Tardieu, 2012). Differences in cardinal temperatures, as well as the relative rates of development within specific temperature ranges, are seen across the thermal functions studied (Table 1).

Despite the importance of thermal functions in the prediction of crop developmental stages and the variation in the currently utilized thermal functions, there is no clear indication

of the relative precision of these functions in simulating crop developmental phases. Fundamental differences in the accuracy of these algorithms are largely unknown, partially due to the use of genotype coefficients that “fit” the genotype to a set of data. Therefore, the evaluation of thermal functions for the accurate simulation of maize development (while limiting potential biases imposed by the use of genotype coefficients) is an important inquiry.

In assessing the impact of climate change on crop productivity, assumptions are generally made about the hybrids adapted for a region or the date on which the hybrid was probably planted. In some applications of crop models, such as gridded models, instead of specific hybrids an “average” hybrid is used to capture the average response of hybrids adapted to the geographic region. To indicate the geographic region of adaptation, commercial hybrids are grouped by comparative relative maturity (Lauer, 1998), or simply relative maturity (RM). The determination of the developmental response of RM groups is therefore of value to modelers to simulate the “average” response of hybrids adapted to specific geographies. Variation in the precision of the thermal functions for different RM groups may indicate that some thermal functions are better able to capture the temperature variation extent in the geographies to which the RM groups are adapted or, alternatively, that RM group hybrids have different temperature responses. Therefore, the relationship between RM groups and thermal function precision is an important consideration. An early vs. a delayed planting date may also impact the precision of thermal function in much the same way as RM group, again calling for consideration of how the precision of thermal functions may vary by the temperature variations that can result as a consequence of a change in planting date.

The overall goal of this study was to bring together researchers to evaluate a number of thermal functions for their ability to predict maize phenology across a range of source data that vary in the number of hybrids, geographies, and environmental conditions tested. The specific objectives of this study were (i) to assess the effect of source data on the ability to differentiate among thermal functions, (ii) to quantify the relative precision of eight thermal functions in predicting maize phenological development, and (iii) to attribute the precision of thermal functions to their response across various temperature ranges.

MATERIALS AND METHODS

Thermal Functions

Eight thermal functions were selected for evaluation and classified according to their derivation type (i.e., empirical or process based) and thermal response. An empirical function is defined herein as a function for which the level of organization of its derivation and parameterization is the same as that of its outcome, e.g., the planting to anthesis interval of a field-grown maize canopy. In contrast, a process-based function is defined as a function that is derived and parameterized at a lower level of organization than that of its outcome, e.g., the derivation and parameterization of a function at the enzyme-kinetics level, whereas the outcome of the function is the planting to anthesis interval of a field-grown maize canopy. For the purposes of this study, the APSIM, TLU, MAIZSIM, and EnzymResp functions (defined below) were classified as process-based functions, and the GDD_{10,30}, CERES-Maize, CHU, and GTI functions (defined

below) were classified as empirical functions. The empirical functions were subdivided into “linear” (GDD_{10,30} and CERES-Maize) and “nonlinear” (CHU and GTI) functions, based on their temperature response at a constant temperature during a 24-h period. The thermal functions evaluated in this study are summarized in Table 1 and defined as follows:

1. GDD_{10,30}: Growing degree days (GDD) with a “base” temperature of 10°C and an “optimum” temperature at 30°C is a thermal function developed by Gilmore and Rogers (1958) using 10 hybrids and 10 inbred lines grown at College Station, TX, at five planting dates in 1956. They calculated heat unit accumulation by subtracting 10°C from the daily minimum and maximum temperatures before calculating daily means, using six combinations of base (none or 10°C) and optimum (none, 30, or 32.2°C) temperatures. The method with the highest precision in their study was the classic 10/30°C cutoff method.
2. CERES-Maize (GDD_{8,34}): The CERES-Maize model utilizes GDD_{8,34} with temperatures at hourly time steps estimated from the minimum and maximum temperatures. The GDD_{8,34} is a linear function similar to GDD_{10,30} but with base and optimum temperatures of 8 and 34°C, respectively. The original thermal function for CERES-Maize described by Jones and Kiniry (1986) used eight 3-h, third-order polynomial interpolations, but in the current version, thermal accumulation is calculated from temperatures during 24 1-h periods (J.T. Ritchie, personal communication, 1998), which are estimated using a sine wave between the minimum and the maximum daily temperatures (cf., Tollenaar et al., 1979). The Hybrid-Maize (Yang et al., 2006) and IXIM (Lizaso et al., 2011) models also use GDD_{8,34} with eight 3-h and 24 1-h temperatures, respectively, estimated from minimum and maximum temperatures.
3. APSIM: The APSIM model uses a single multilinear function for thermal accumulation that reflects the biological response of development across the 0 to 44°C range in temperatures encountered in most environments in which maize is cultivated (Wilson et al., 1995). This temperature-response function has 0°C minimum, 34°C optimum, and 44°C ceiling temperatures. Daily thermal accumulation was calculated from eight 3-h, third-order polynomial interpolations between the minimum and maximum daily temperatures.
4. Thermal leaf units (TLU). Analogous to accumulated growing degrees days in GDD_{10,30} (°C d), TLU is accumulated thermal leaf units (L, leaf tips) by conversion of temperature to leaf (tip) units through the temperature-dependent rate of leaf appearance ($L \text{ } ^\circ\text{C}^{-1} \text{ d}^{-1} \times \text{ } ^\circ\text{C d} = L$), where the rate of leaf appearance (RLA) is the inverse of the phylochron. The relationship between the rate of leaf-tip appearance and temperature (Tollenaar et al., 1979) represents a biologically meaningful temperature response. This response is similar to other growth and development processes in maize such as seedling growth (Lehenbauer, 1914), radical elongation (Blacklow, 1972), and leaf elongation (Parent and Tardieu, 2012). The temperature-compensated rate of leaf-tip appearance is fairly constant until the emergence of the topmost leaf (Tollenaar et al., 1979, 1984). The number of visible leaf tips per plant is assumed to be equal to the accumulated TLU at any time between planting and the appearance of

the topmost leaf if the rate of leaf appearance is influenced by temperature only. Based on this assumption, TLU accumulation between planting and anthesis is equal to the total leaf number plus the TLU accumulation during the interval between the appearance of the topmost leaf and anthesis. Leaf-ligulae appearance has also been used to quantify (e.g., Warrington and Kanemasu, 1983a) and to model (e.g., Parent et al., 2010) the rate of leaf appearance and maize development, among others, because of the wide use of V stages (Abendroth et al., 2011) to describe phenological development in maize. In contrast to the temperature-compensated rate of leaf-tip appearance, the temperature-compensated rate of leaf-ligule appearance varies widely for the planting to flowering interval. The ratio of the rate of leaf-tip appearance to the rate of leaf-ligule appearance is >1 during the first eight to 12 leaves, and it is <1 during the second half of the pre-silking period (Muldoon et al., 1984). The variability of the temperature-compensated rate of leaf-ligule appearance across the pre-silking phase diminishes its suitability for use in modeling the rate of maize development. Analogous to accumulated growing degrees days in $GDD_{10,30}$ ($^{\circ}C d$), TLU, ($L^{\circ}C^{-1} d^{-1} \times ^{\circ}C d = L$) are accumulated by the conversion of temperature to leaf tips through the RLA function.

5. MAZSIM (β function): A β function was introduced by Yin et al. (1995) that described the temperature response of crop development. This function describes a smooth non-symmetric response to temperature using five parameters: three cardinal temperatures (the base temperature [T_{base}], the optimum temperature [T_{opt}], and the maximum temperature [T_{ceil}]), a parameter for the maximum rate of development at T_{opt} (R_{max}), and a parameter that describes the curvature of the relationship. All parameters except the parameter that describes the curvature are biologically meaningful. Yan and Hunt (1999) simplified the Yin et al. (1995) function by eliminating the variable describing the shape of the curve and by setting T_{base} to zero, resulting in the following equation with three parameters:

$$R(T) = R_{max} \left(\frac{T_{ceil} - T}{T_{ceil} - T_{opt}} \right) \left(\frac{T}{T_{opt}} \right)^{T_{opt}/(T_{ceil} - T_{opt})} \quad [1]$$

where $R(T)$ is the rate of development as a function of temperature. Yan and Hunt (1999) showed that this simplified β function was highly predictive when model parameters that were estimated from six constant temperatures were used to predict the rates of 16 varying day-night temperature regimes in the Tollenaar et al. (1979) data set. The MAZSIM model (Kim et al., 2012) uses Eq. [1] for predicting the leaf appearance rate and assigned the following parameter values: $T_{opt} = 31.2^{\circ}C$, $T_{ceil} = 43.7^{\circ}C$, and $R_{max} = 0.53 d^{-1}$.

6. Enzymatic response (EnzymResp): Using an equation for describing the temperature response of enzyme activities (Johnson et al., 1942), Parent et al. (2010) showed that processes linked to plant growth and development showed coordinated temperature responses and followed common laws within a plant species. Interestingly, they showed that there was no coordination of temperature responses linked to plant metabolism and leaf photosynthesis. In a meta-

analysis, Parent and Tardieu (2012) utilized an equation to show a common response among maize genotypes from high-latitude, temperate, and tropical regions across a wide range in temperatures:

$$R(T) = \frac{AT \exp(-\Delta H_A/RT)}{1 + [\exp(-\Delta H_A/RT)]^{\alpha(1-T/T_0)}} \quad [2]$$

The function has two parameters: $T_0 = 306.4^{\circ}K (= 33.3^{\circ}C)$ and $\alpha = 3.5$ (Parent and Tardieu, 2012). The parameter $\Delta H_A = 73900 J mol^{-1}$ is deduced from the other two, and A is a size factor without units (51,559,240,052).

7. Crop heat units (CHU): Crop heat units have been used in Canada to account for temperature effects on phenology in maize since the mid-1960s. Crop heat units are estimated using separate night and day temperature functions (Table 1). The former is a linear function with a minimum of $4.4^{\circ}C$ and the latter is a quadratic function with a minimum at $10^{\circ}C$ and an optimum near $30^{\circ}C$ (Brown, 1969). The quadratic function was developed by Brown (1960) from data collected on soybean [*Glycine max* (L.) Merr.] grown in environment-controlled studies (Van Schaik and Probst, 1958).
8. General thermal index (GTI): The GTI model is based on a large data set of maize hybrids grown during 4 yr at 19 locations across a 9° interval in latitude across the North American Corn Belt, in which the durations from planting to silking and from silking to maturity were related to air temperature (Stewart et al., 1998). The GTI model is unique in that (i) it consists of two polynomial responses to temperature, one for the period from planting to silking and one for the period from silking to maturity, and (ii) the temperature response during the post-silking period was flat relative to that during the pre-silking period, particularly for temperatures $<20^{\circ}C$ (cf., Stewart et al., 1998, Fig. 2 and 5). Although the relative insensitivity of the duration of the grain-filling period in maize to temperature has been reported previously (Shaw and Thom, 1951; Brown, 1977), none of the current thermal models other than GTI account for differences in the temperature response between the pre- and post-flowering periods.

Thermal functions can have a range of responses across temperatures and, consequently, the precision of the thermal functions will probably be related to their differential response across the temperature ranges to which they are exposed. Most figures of thermal functions in the literature graph the developmental response against a specific temperature value, which is useful for evaluating the function at the hour when the plant experienced the specific temperature. However, due to the diurnal range in temperatures to which plants are exposed daily, an hour snap shot is a limited perspective of the plant's daily developmental response. Given that a location's diurnal temperature variation is relatively stable, the response of thermal functions for a given diurnal range can be computed and graphed against a range of mean temperatures. To better capture how the thermal functions in this study respond to daily ambient conditions, the response of the thermal functions to mean temperature in the 0 to $40^{\circ}C$ range was calculated assuming a diurnal temperature range of $12^{\circ}C$ (i.e., the difference between T_{max} and T_{min} values). The $12^{\circ}C$ diurnal

temperature range value was selected because this was the approximate mean diurnal temperature range in the data sets utilized in the current study. To facilitate a comparison of the relative response, each function was normalized for the rate at its optimum temperature (Fig. 1).

Data Sources and Analyses

The precision of the thermal functions was evaluated using data from three different sources:

1. North America: Data were collected in field trials performed by Monsanto Co. that resulted in two different data sets. First, data were collected in trials that were performed from 2007 to 2011 at 43 locations across the North American Corn Belt. A total of 118 commercial DeKalb maize hybrids were tested that ranged from 76 to 119 RM. The hybrids were grown in their area of adaptation, and the number of locations in each of five RM classes was 13 for RM 76 to 85, 27 for RM 86 to 95, 26 for RM 96 to 105, 28 for RM 106 to 115, and 14 for RM 116 to 119 (i.e., the total number of locations is 43 because hybrids of more than one RM class were tested in several locations). The number of hybrids and locations tested varied each year. The data set consisted of 1375 combinations of hybrids \times locations \times years. Dates of planting and 50% anthesis were measured in this data set. The second data set was collected in trials that were performed from 2007 to 2012 at a location near DeKalb, IL. This location was not included in the 43-location North American data set. At this location, dates of black layer formation (i.e., physiological maturity), anthesis, and silking were recorded on 3129 hybrids \times years, and the hybrids tested included all 118 hybrids grown at the other 43 locations. The hybrids in this data set ranged in RM from 76 to 119. At all locations, maize hybrids were grown in 6.1-m-long, four-row plots using standard agronomic practices.
2. Indiana–Ohio: This data set consisted of 108 entries. Details of this study have been previously reported by Nielsen et al. (2002). In short, three Pioneer maize hybrids (CRM 106, 111, and 115) were grown at three planting dates, ranging from 22 April to 17 June, each at four locations in Ohio and Indiana from 1991 to 1994. Note that Pioneer uses CRM (comparative relative maturity) to designate hybrid RM and that hybrid RM ratings may vary somewhat among different seed companies (Lauer, 1998). Dates of 50% silking and 50% black layer were recorded.
3. Argentina: This data set consists of information collected on various maize hybrids ranging from RM 92 and 127 that were grown between 1988 and 2013 near Balcarce, Argentina. Data included the results of a published study (Cirilo and Andrade, 1994a, 1994b) and several unpublished studies. Some of the studies had multiple planting dates that ranged from September to January. The experimental area was irrigated when deemed necessary. Dates of 50% silking and 50% black layer were recorded.

Weather data were collected for each location-year in the three data sets. Records of daily maximum and minimum air temperatures were obtained either from the National Climate Data Center (U.S. weather stations, <http://www.ncdc.noaa.gov/cdo-web/search>) or the Government of Canada Climate Service (Canadian weather stations,

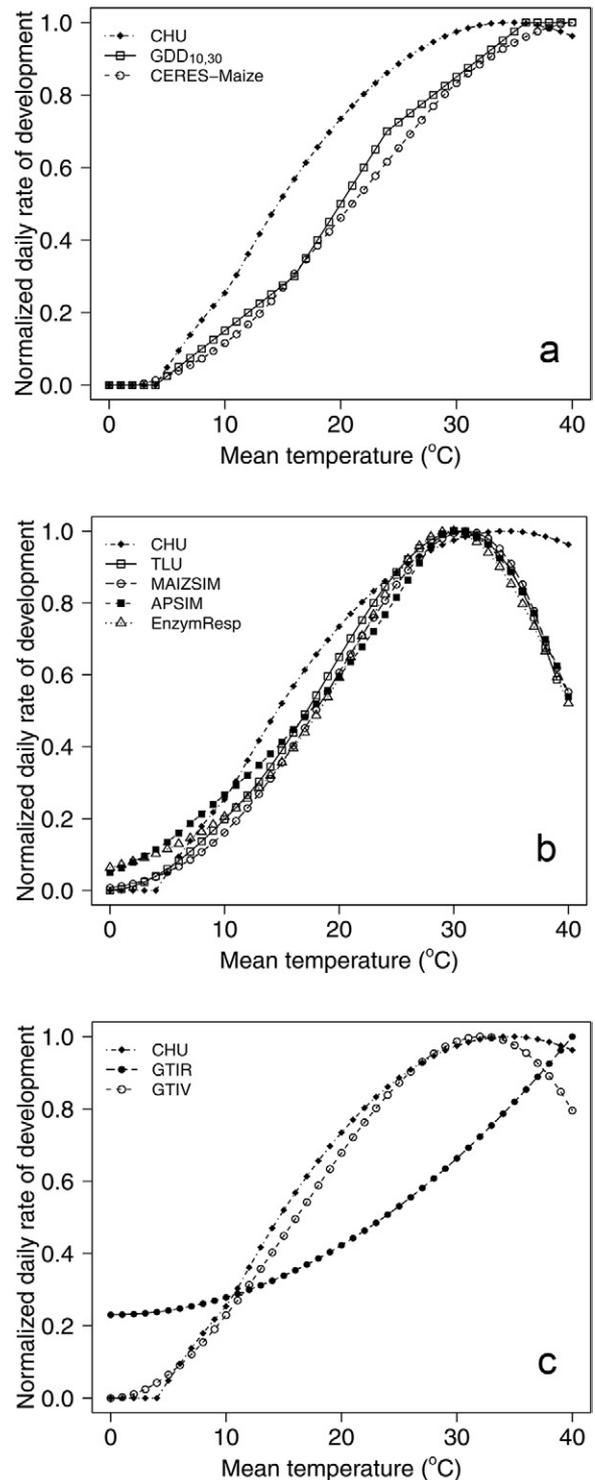


Fig. 1. Relationships between mean daily temperatures with a 12°C diurnal range normalized for rate of development at the optimum temperature specific to each function: (a) growing degree days (GDD_{10,30}), CERES-Maize, and crop heat units (CHU) functions, (b) APSIM, thermal leaf units (TLU), MAZSIM, EnzymResp, and CHU functions, and (c) general thermal index (GTI) during the preflowering period (GTIV), GTI during the post-flowering period (GTIR), and CHU functions.

http://climate.weather.gc.ca/index_e.html#access) for the location-years in the North American and Indiana–Ohio data sets. All weather service stations were, on average, within 20 km of the experimental site.

Field data were analyzed for the pre-flowering and post-flowering periods. The pre-flowering period was defined as the period from the first day after planting until, and including, anthesis (i.e., the first day at which $\geq 50\%$ of plants shed pollen) or silking (i.e., the first day at which silks had emerged from the topmost ear shoot in $\geq 50\%$ of plants), and the post-silking period was defined as the first day after silking until, and including, black layer (the first day on which 50% of plants have grain with a black layer in the center portion of the ear). Whenever possible, the planting to anthesis interval rather than the planting to silking interval was analyzed because the anthesis to silking interval is highly affected by abiotic stresses (Edmeades et al., 2000) and, consequently, the use of the silking date will introduce another potential unaccounted source of variability in the data.

Daily thermal accumulation was estimated using the daily mean temperature (GTI), minimum and maximum temperatures (GDD_{10,30} and CHU), 1-h temperatures (CERES-Maize, TLU, MAZSIM, and EnzymResp), and 3-h temperatures (APSIM). The 1-h temperatures were estimated using a sine wave between the minimum and the maximum daily temperature (e.g., Tollenaar et al., 1979). The 3-h temperatures in the APSIM function were estimated using range fractions for each of the eight time periods within a 24-h day that describe a sine wave.

The CV (coefficient of variability) was used to quantify the precision of the thermal functions. The mean thermal accumulation (and days), standard deviation, and CV were calculated from all observations (i.e., hybrid-location-year) within an RM class for the North American data set and within a planting date for the Indiana–Ohio and Argentinean data sets for the planting to anthesis (or silking) interval and the silking to physiological maturity interval. Statistical analyses of the CVs were performed using the Glimmix procedure for generalized linear modeling assuming a log link and γ distribution in SAS, Version 9.3, where RM classes or planting dates were replications for the statistical comparisons among thermal functions, and thermal functions were replications for the statistical comparisons among either RM classes or planting dates.

Differences in the precision of the thermal functions across RM groups may be due to genetic or environmental factors associated with the regions to which the RM groups are adapted. One method for deconstructing these factors is to identify differences in cardinal temperatures among RM classes. Since the MAZSIM function explicitly quantifies T_{opt} and T_{ceil} values, optimization of this function for each of the five RM classes can identify these cardinal temperatures of each RM class. The optimization procedure of the β function was performed by varying the values of T_{opt} and T_{ceil} (keeping T_{base} unchanged at 0°C) and selection based on the lowest CV. The optimization of the β function was done using the optimum function of the R statistical language (R Development Core Team, 2013) with the Nelder–Mead algorithm for both entries in each of the five RM classes of the 43-location North American data set and for all RM classes combined (β_{OPT}). This is a general purpose routine for function error minimization. Several of the minimization calculations were

run for two different starting values, and the results were identical in the two cases. This exercise confirmed that the algorithm was converging to a global rather than to a local optimum.

In addition, for each of the five RM class in the 43-location North America data set, the hybrid-location-years of the data set were subdivided equally into three groups representing 33% low, 33% medium, and 33% high means of daily minimum temperature and of daily maximum temperature, resulting in 15 mean-minimum and 15 mean-maximum temperature groups (i.e., 5 RM classes \times 3 groups within each RM class). This analysis is informative of thermal function sensitivity variation by temperature. The thermal functions tested vary in T_{base} , T_{opt} , and T_{max} values. Differences in cardinal temperature may render some thermal functions more sensitive to specific temperatures than others, thus impacting their relative precision in simulating development.

RESULTS AND DISCUSSION

Impact of Data Source on Thermal Function Evaluation

Data source is an important variable when evaluating the precision of thermal functions. The data sets evaluated in this study each had limitations. The Indiana–Ohio data set was comprised of a multilocation, multiyear study with orthogonal treatments, but contained only a relatively small number of hybrids and environments (i.e., location-years). In contrast, the North American data sets consisted of a large number of hybrids and environments, but the treatments were not orthogonal. Different hybrids were grown in each location-year, resulting in a different number of observations for each RM class in the 43-location data set (Tables 2 and 3). In addition, the silking to black layer interval was evaluated only at a single geographic location near DeKalb, IL, for multiple years (Table 3). The Argentinean data set was not orthogonal and contained a relatively small number of environments (years).

Empirical nonlinear functions were more precise than either empirical linear or process-based functions in the North American and Indiana–Ohio data sets, but the precision of the thermal functions did not differ in the Argentinean data set (Tables 2–6). Differences among thermal functions and calendar days were generally smaller in the Indiana–Ohio than the North American data set. The precision of the thermal functions for the planting to anthesis phase in the North American data set were calendar days < empirical linear < process based < empirical nonlinear, whereas in the Indiana–Ohio data set they were calendar days < empirical linear = process based < empirical nonlinear. In the Argentinean data set, calendar days were less precise than the thermal functions, but the thermal functions could not be differentiated. Therefore, large data sets have improved ability to differentiate thermal function performance, even when they may have limitations such as lack of orthogonality. Ideally, large and orthogonal data sets would be preferred.

Relative Precision of Thermal Functions

Within the large data sets that allowed the greatest differentiation among the thermal functions, the empirical nonlinear functions proved to be more precise than the other functions tested (Tables 2–6). Thermal functions were generally more precise than calendar days, although differences were

Table 2. Coefficients of variation of the number of days and thermal accumulation during the planting to anthesis interval determined by eight functions for commercial hybrids across five relative maturity (RM) groups grown at 43 locations in the Corn Belt from 2007 to 2011 (North American data set). The number of observations (*n*) is the sum of all hybrids grown in each location-year for a RM class.

RM	<i>n</i>	Coefficient of variation								
		Days	GDD _{10,30}	CERES	MAIZSIM	TLU	EnzymResp	APSIM	GTI	CHU
		%								
76–85	121	10.3	9.5	9.3	9.0	8.4	9.0	7.8	7.6	7.0
86–95	204	9.9	9.7	9.5	9.2	8.5	9.2	7.7	7.6	6.8
96–105	342	9.3	6.4	6.5	6.2	6.0	6.3	6.0	5.7	5.6
106–115	560	10.4	6.4	6.9	6.0	5.3	6.2	5.4	4.7	4.5
116–122	148	10.3	6.3	6.9	5.9	5.3	6.0	5.2	4.5	4.6
Mean		10.1 e†	7.7 d	7.8 d	7.3 cd	6.7 c	7.3 d	6.4 bc	6.0 ab	5.7 a
All	1375	10.7	10.1	10.2	9.5	8.7	9.6	7.9	7.5	6.7

† Means within a row followed by the same letter are not significantly different at the 0.05 level of probability.

Table 3. Coefficients of variation of the number of days and thermal accumulation during the silking to black layer interval determined by eight thermal functions for commercial hybrids across five relative maturity (RM) classes grown at a single location in the U.S. Corn Belt from 2007 to 2012 (North American data set). The number of observations (*n*) is the sum of all hybrids grown in each location-year for a RM class.

RM	<i>n</i>	Coefficient of variation								
		Days	GDD _{10,30}	CERES	MAIZSIM	TLU	EnzymResp	APSIM	GTI	CHU
		%								
76–85	182	9.0	8.2	8.0	7.6	7.2	8.2	7.1	6.2	6.0
86–95	413	9.4	9.3	9.0	8.6	8.2	9.3	8.2	7.4	6.8
96–105	771	8.5	9.4	9.3	8.7	8.0	9.5	8.3	7.1	6.0
106–115	1477	9.9	9.6	9.4	8.9	8.2	9.6	8.1	7.1	5.8
116–122	286	9.6	11.8	11.5	11.0	10.3	11.3	9.6	7.7	7.4
Mean		9.3 de†	9.7 e	9.5 de	9.0 d	8.4 c	9.6 e	8.3 c	7.1 b	6.4 a
All	3129	12.4	10.2	10.1	9.7	9.1	10.3	9.4	9.0	7.7

† Means within a row followed by the same letter are not significantly different at the 0.05 level of probability.

Table 4. Coefficients of variation of the number of days and thermal accumulation during the planting to silking and silking to black layer intervals determined by eight thermal functions for three maize hybrids planted at three dates from April to June during 1990 to 1994 at four locations in Ohio and Indiana (Nielsen et al., 2002); the total number of observations is 108. Data were analyzed both separately for the early, medium, and late planting dates and together for all planting dates.

Planting date	Days	Coefficient of variation								
		GDD _{10,30}	CERES	MAIZSIM	TLU	EnzymResp	APSIM	GTI	CHU	
		%								
		<u>Planting to silking</u>								
Early	11.6	4.4	5.2	4.7	3.6	4.5	5.9	4.4	3.5	
Medium	9.1	4.7	5.4	4.9	3.9	4.9	6.1	4.5	3.8	
Late	5.6	5.7	6.4	6.1	5.2	6.1	7.2	5.7	4.7	
Mean	8.8e†	4.9abc	5.7cd	5.3bcd	4.2ab	5.2bcd	6.4d	4.8abc	4.0a	
All	12.9	5.2	5.8	5.5	4.7	5.5	7.0	5.4	5.0	
		<u>Silking to black layer</u>								
Early	9.5	8.4	9.2	9.0	7.9	8.7	10.1	6.8	8.0	
Medium	11.5	9.0	10.2	10.0	8.6	9.4	11.0	7.6	8.9	
Late	8.6	11.3	12.7	12.5	10.7	11.8	14.0	8.6	10.5	
Mean	9.9bc	9.6bc	10.7cd	10.5cd	9.0b	10.0bc	11.7d	7.7a	9.1b	
All	10.3	11.8	13.1	12.8	11.0	12.0	12.8	7.9	10.5	

† Means within row followed by the same letter are not significantly different at 0.05 level of probability.

smaller during the period between silking and black layer than during the planting to anthesis interval. The precision of the thermal functions for both the pre-flowering and post-flowering period in the North American data set was empirical linear < process based < empirical nonlinear (Table 6). The CHU and GTI functions were the most precise for the prediction of both the pre-anthesis and post-silking phases in the North American data set (Tables 2 and 3). In the Indiana–Ohio data set, the GTI function was the only thermal function that was more precise than calendar days for the silking to black layer period (Table 4). The post-flowering GTI thermal function had a very different

thermal response than any other thermal function tested (Fig. 1). The higher precision of GTI for the post-flowering period was associated with a relatively small difference in thermal accumulation from silking to maturity between the early and late planting dates for GTI. Nielsen et al. (2002) showed a 17% difference in GDD_{10,30} accumulation during the post-flowering period between early and late planting, whereas the difference in GTI accumulation was only 5% (data not shown). The mean minimum daily temperatures during the silking to black layer interval decreased from 14.3 to 10.7°C for the early and late planting dates, respectively.

Table 5. Coefficients of variation of number of days and thermal accumulation by eight functions during the planting-silking and silking-black layer intervals of maize hybrids planted from September to January between 1989 and 2012 at locations near Balcarce (Argentina). Data were analyzed both separately for four planting-date periods and together for all planting dates. The number of observations (*n*) is the sum of all hybrids grown in each location-year for a RM class.

Planting date	<i>n</i>	Coefficient of variation								
		Days	GDD _{10,30}	CERES	MAIZSIM	TLU	EnzymResp	APSIM	GTI	CHU
%										
Planting to silking										
Sept.	6	5.4	8.2	7.8	7.7	7.6	7.6	6.7	7.0	6.9
Oct.	41	6.7	6.2	6.5	6.2	5.6	6.4	6.5	6.3	5.5
Nov.	13	9.0	8.5	8.7	8.5	8.3	8.7	8.6	8.4	8.1
Dec.–Jan.	16	12.5	9.9	10.0	9.9	10.0	9.9	10.2	10.2	10.8
Mean		8.4 a†	8.2 a	8.2 a	8.1 a	7.9 a	8.2 a	8.0 a	8.0 a	7.8 a
All	76	18.5	8.8	9.1	9.0	9.0	9.2	10.3	10.1	10.7
Silking to black layer										
Sept.	6	6.7	6.0	5.0	5.4	5.7	5.6	5.1	5.4	5.4
Oct.	41	12.5	12.6	13.0	12.7	12.1	12.8	12.9	12.7	11.9
Nov.	13	18.7	11.0	10.3	10.8	11.9	11.0	11.8	13.6	13.3
Dec.–Jan.	16	11.0	8.6	8.5	8.4	8.4	8.1	7.6	8.2	8.7
Mean		12.2 b	9.6 a	9.2 a	9.3 a	9.5 a	9.4 a	9.4 a	10.0 a	9.9 a
All	76	13.0	17.0	17.5	16.8	15.7	17.0	15.7	14.1	14.0

† Means within row followed by the same letter are not significantly different at 0.05 level of probability.

Table 6. Mean coefficients of variation of calendar days and thermal accumulation determined by empirical-linear functions (GDD_{10,30} and CERES-Maize), process-based functions (APSIM, MAIZSIM, EnzymResp, and TLU), and empirical-nonlinear functions (GTI and CHU) during the planting to anthesis or silking and the silking to black layer intervals in the 43-location, 5-yr and the one-location, 6-yr North American data sets (Tables 2 and 3), the Indiana–Ohio data set (Table 4), and the Argentinean data set (Table 5).

Phase of development	Data set	Coefficient of variation			
		Calendar days	Thermal function		
			Empirical linear	Process-based	Empirical nonlinear
%					
Planting to anthesis or silking	North America	10.1 d†	7.7 c	6.9 b	5.9 a
	Indiana–Ohio	8.8 c	5.3 b	5.3 b	4.4 a
	Argentina	8.4	8.2	8.0	7.9
Silking to black layer	North America	9.3 c	9.6 c	8.8 b	6.8 a
	Indiana–Ohio	9.9 ab	10.1 b	10.3 b	8.4 a
	Argentina	12.2 b	9.4 a	9.4 a	9.8 a

† Means within a row followed by the same letter are not significantly different at the 0.05 level of probability.

Relationship between Temperature Response and Precision of Thermal Functions

The precision of the thermal functions is associated with their response across a wide range of temperatures. One means of understanding the precision of thermal functions is to compare their normalized response across a range of temperatures in relation to that of CHU, which has been shown to be more precise in the North American data set (Fig. 1). The results of the normalized responses to mean temperatures (based on a 12°C diurnal range) show that: (i) the empirical functions (GDD_{10,30} and CERES) have a linear-like response between mean temperatures of 4 and 36°C (Fig. 1a) (the generally depicted temperature response of GDD_{10,30} in the literature is a straight line between 10 and 30°C, but this is true only when there is no diurnal temperature variation or if the maximum and minimum daily temperatures are >10°C and <30°C); (ii) the process-based functions (TLU, APSIM, MAIZSIM, and EnzymResp) have a temperature response similar to each other in the 15 to 45°C temperature range, with small differences in the lower temperature range (Fig. 1b); and (iii) the nonlinear empirical functions are similar to each other across 0 to 30°C for the pre-silking phase (Fig. 1c). The post-silking GTI function has a very

different temperature response that is relatively insensitive at temperatures <23°C (Stewart et al., 1998).

In this study, the nonlinear empirical functions were the most precise group of thermal functions tested, and differences in precision were associated with response in the 10 to 26°C mean diurnal temperature range (Fig. 1). Unlike linear empirical functions, the shape of process-based functions and the nonlinear empirical function are similar (Fig. 1). These two groups of thermal functions varied somewhat in their base temperatures (Fig. 1b and 1c), but there was no consistent relationship between the base temperatures and the precision of the function (Tables 2–5). All three groups differ in the supra-optimal temperature range. However, differences in precision cannot be attributed to the developmental responses in the supra-optimal temperature range because none of the data sets used in this study contained mean daily temperatures >30°C. Differences in precision among thermal functions were associated with their relative temperature response in the 10 to 26°C mean diurnal temperature range (Fig. 1). The CHU function had higher developmental rates across the 10 to 26°C range and was the most precise in predicting phenology. At 20°C, for instance, the rate of development of CHU is 21% greater than that of TLU, and the rate of development of TLU is 21% greater than that of GDD_{10,30} (Fig.

Table 7. Mean coefficient of variations (CV) of thermal accumulation determined by eight functions during the planting to anthesis and silking to black layer intervals, and mean daily minimum and maximum temperatures of locations for five relative maturity (RM) groups. The planting-to-anthesis interval comprised observations across 43 Corn-Belt locations during 2007 to 2011 (Table 1) and the silking to black layer interval comprised > 3100 observations at one Corn-Belt location from 2007 to 2012 (North American data sets).

RM	CV	Planting to anthesis		CV	Silking to black layer	
		Daily temperature			Daily temperature	
		Min.	Max.		Min.	Max.
d	%	°C		%	°C	
76–85	8.5 c†	11.7	23.1	7.3 a	17.1	28.4
86–95	8.5 c	12.1	23.9	8.4 b	16.9	27.9
96–105	6.1 b	13.0	25.4	8.3 b	16.6	27.7
106–115	5.7 a	14.3	26.3	8.3 b	15.6	26.7
116–122	5.6 a	15.7	27.5	10.1 c	15.1	26.3

† Means within a column followed by the same letter are not significantly different at the 0.05 level of probability.

1), while precision of the three functions in the North American data sets was in the order CHU > TLU > GDD_{10,30} (Tables 2 and 3). Thermal functions with higher normalized developmental rates across a wide range of temperatures (e.g., nonlinear empirical functions) have lower temperature sensitivity.

The precision of the thermal functions varied with RM class in the North American data set, and the variation in precision was associated with air temperature. The precision of the thermal functions increased from low- to high-RM classes during the planting to anthesis interval (Table 7) in the 43-location North America data set. This relationship between RM class and thermal function precision could be due to either genetics or environment. The association with genetics was dismissed because, when all RM hybrids were grown at a single location (i.e., the DeKalb, IL location), the trend in RM precision was no longer observed, i.e., there was either an opposite trend during the silking to black layer period (Table 7) or no trend during the planting to anthesis interval (data not shown). The effects of temperature on the relative precision of thermal methods across the RM groups in the North American data set was examined in more detail by subdividing each of the five RM groups of environments into three mean-minimum-temperature environments (Fig. 2). In this analysis, the change in CV of each thermal function across the 15 temperature–RM groups

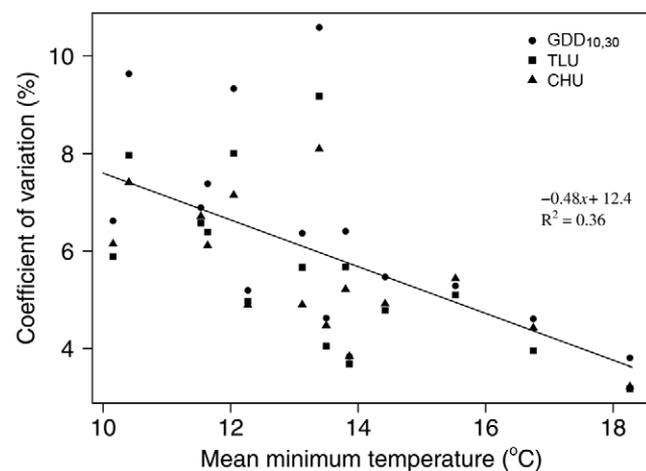


Fig. 2. Relationship between mean minimum temperature and CVs of the growing degree day (GDD_{10,30}), thermal leaf units (TLU), and crop heat units (CHU) functions for 12 environments (location-years). Environments were grouped into the 33% highest, 33% medium, and 33% lowest mean minimum temperature environments in each of the five relative maturity classes depicted in Table 7.

was $-0.55\% \text{ } ^\circ\text{C}^{-1}$ for GDD_{10,30}, $-0.49\% \text{ } ^\circ\text{C}^{-1}$ for TLU, and $-0.41\% \text{ } ^\circ\text{C}^{-1}$ for CHU, but these slopes were not significantly different (data not shown). The CVs in some of the temperature environments deviated substantially from the regression (Fig. 2), and variation among the three thermal functions within these environments was small when the deviation from the regression was negative (e.g., the 10.2, 12.3, and 13.9°C environments) and was large when the deviation from the regression was positive (e.g., the 10.4, 12.0, and 13.4°C environments).

When the β function was optimized for five RM groups, the values of T_{opt} and T_{ceil} of the early RM classes (Table 8) were much lower than reported values from controlled-environment studies (e.g., Parent et al., 2010; Sanchez et al., 2014). The low T_{opt} and T_{ceil} values were probably not due to genetic differences among RM groups. Low T_{opt} (21.2 and 22.3°C) has been reported for the temperature-dependent duration of the period from sowing to tassel initiation for two highland tropical inbred lines in controlled-environment studies performed by Ellis et al. (1992) and reported by Yin et al. (1995), but neither the results of Tollenaar et al. (1979), which included North American hybrids of <85 RM, nor the results of Parent and Tardieu (2012), which included very early European hybrids and inbred lines, showed T_{opt} and T_{ceil} values that were lower than those reported in studies of temperature-dependent rates of growth and development in maize (cf., Parent and Tardieu, 2012).

The relatively high precision of temperature-insensitive functions, the decline in CV with increase in temperature among RM groups, and low cardinal temperatures in the optimized β function of early RM groups all support the contention that maize phenology is influenced by factors other than air temperature per se. Factors other than air temperature

Table 8. Coefficients of variation (CV) for the planting to anthesis interval of thermal functions that utilize different forms of the β function: the MAIZSIM β function, and different β functions modified in order to optimize CV by changing T_{opt} and T_{ceil} within each of the five relative maturity (RM) classes (T_{base} is held constant at 0°C).

RM	Coefficient of variation	
	β_{OPT}	MAIZSIM
d	%	
76–85	6.2 (23.0, 32.4)†	9.0 (32.1, 43.7)
86–95	6.3 (25.7, 39.6)	9.2 (32.1, 43.7)
96–105	5.4 (26.2, 35.9)	6.2 (32.1, 43.7)
106–115	4.3 (27.6, 37.8)	6.0 (32.1, 43.7)
116–122	4.3 (30.3, 44.9)	5.9 (32.1, 43.7)

† T_{opt} , T_{ceil} (°C) in parentheses.

that can influence phenology include soil and apex temperature (Vinocur and Ritchie, 2001), photoperiod (e.g., Kiniry et al., 1983; Tollenaar and Hunter, 1983; Warrington and Kanemasu, 1983b), incident solar radiation (Birch et al., 1998, Tollenaar, 1999), and periods of severe stress (e.g., McCullough et al., 1994). For instance, apex temperature is influenced by plant transpiration and soil temperature before the eight-leaf-tip stage and the apex-air temperature differential appears to be greatest at low temperatures (Vinocur and Ritchie, 2001; Birch et al., 1998). The apex-air temperature differential may have played a role in the higher precision of less temperature-sensitive functions, the increase in precision with temperature among RM groups, and the low T_{opt} and T_{ceil} values for the early RM groups in the MAIZSIM optimization. The lower precision of process-based functions may be due to the fact that these data sets were generated under controlled-environment conditions where many of the factors other than air temperature were held constant and where the apex-air temperature differential may have been less than under field conditions.

Precision of Thermal Functions under Supra-Optimal Temperatures

Climate change will probably result in an increase in the number of days in the growing season with elevated temperatures, and therefore an accurate estimate of the impact of supra-optimal temperatures on maize phenology will be important in evaluating the impact of climate change on maize yield. The response to supra-optimal temperature differs substantially among the thermal functions (Fig. 1). The process-based thermal functions TLU, MAIZSIM, and EnzymResp are derived from data that includes supra-optimal temperatures. These functions were derived from controlled-environment studies during early phases of development that showed that the rate of development declines rapidly when temperatures are raised beyond the optimum temperature (Lehenbauer, 1914; Tollenaar et al., 1979; Warrington and Kanemasu, 1983a,b; Parent and Tardieu, 2012). In contrast, the rate of development remains constant beyond 36°C for empirical linear functions and declines moderately beyond the optimum temperature in nonlinear empirical functions that account for the whole life cycle (Fig. 1). Nonlinear empirical functions were derived based on data sets that contained either no or few supra-optimal temperatures (i.e., Van Schaik and Probst, 1958; Brown, 1960; Stewart et al., 1998), and the precision of these functions may be low for supra-optimal conditions. Hence, while data are limited on the supra-optimal range, we suggest that maize models that are used to assess the impact of climate change on crop productivity should utilize the developmental response to supra-optimal temperatures of process-based functions for the pre-flowering period. Even less is known about the developmental response to supra-optimal temperatures during the grain-filling period. The GTI and CHU functions performed best during the post-silking period in the data sets evaluated here (Tables 3 and 4), with mean temperatures <25°C. To the best of our knowledge, no information is available on the response of the grain-filling period to supra-optimal temperatures. Owing to the paucity of supra-optimal temperature data in the derivation or evaluation of empirical thermal functions, process-based functions appear to better represent the temperature response of maize development under supra-optimal temperatures. Overestimation of the rate of development under supra-optimal

temperatures by empirical models will lead to a disproportionate reduction of the life cycle, and consequently, models that use empirical thermal functions will underpredict yield in climate-change scenarios.

CONCLUSIONS

The results of this study show that large data sets can improve the ability to differentiate among thermal functions that quantify the effect of temperature on maize phenology. The precision of nonlinear empirical functions, particularly CHU, is superior to that of both linear empirical functions, such as GDD_{10,30}, and process-based functions, such as the TLU function, in predicting maize phenology. The precision of thermal functions was associated with the temperature response across the 10 to 26°C temperature range rather than with the base and optimum cardinal temperatures or the supra-optimal temperature range. The higher precision of the CHU function is associated with its overall lower relative temperature sensitivity in the 10 to 26°C temperature range, which is probably due to factors other than air temperature that influence rate of development. The GTI function during the post-silking period has a low temperature sensitivity for temperatures <23°C (Fig. 1c), and this function was superior for the Indiana–Ohio data set. Because the nonlinear empirical functions were developed under conditions devoid of supra-optimal temperatures, and while we await for research addressing the response to temperature in that range, maize models that are used to assess the impact of climate change on crop productivity should utilize the developmental response of process-based functions to quantify the response of phenology to supra-optimal temperatures during the pre-flowering period.

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